everest biolabs

Atlas
HSA ELISA Quick Guide

Revision 1.3 – December 10, 2025

The Atlas HSA ELISA is an immunoassay designed to measure Human Serum Albumin (HSA) in plasma, serum, urine, and cerebrospinal fluid (CSF). This kit contains enough reagents for 96 wells. This is a high-sensitivity assay, and it is critical to avoid contamination for reliable results. Always clean the work area, use fresh tips, and avoid contamination of buffers.

Supplied kit contents:

Item	Quantity	Storage
HSA Capture Antibody 100X	80 µL	4° C
HSA Detection Antibody 100X	80 µL	4° C
Diluent	50 mL	4° C
Wash Buffer 10X	25 mL	4° C or Room temperature (RT)
TMB Solution	14 mL	4° C
Stop Solution	14 mL	4° C or RT
Pre-Coated 96-Well Microplate	96-well	4° C
Plate Seals	3	RT
HSA standard	5 x 0.2 µg	4° C or -20° C

Materials needed:

- Microplate reader (450 nm absorbance)
- Plate Shaker
- Low-bind Eppendorf tubes
- Lab-grade deionized water
- 8 channel pipettor (300 µL) or a plate washer (recommended)

About the kit:

- The kit provides enough reagents for 96 wells (12 x 8 well strips).
- Capture and detection antibodies have only been tested for stability in the 100X formulations.
- **This assay is for research use only, not for diagnostic procedures.**

Reagent preparation:

- Equilibrate all reagents to room temperature (18-25°C) before use.
- Briefly spin down capture and detection antibody stocks.
- The volumes below are sufficient for one strip (8 wells); adjust volumes as needed.
- Prepare only the required volume of reagents for each experiment.

1X Wash Buffer:

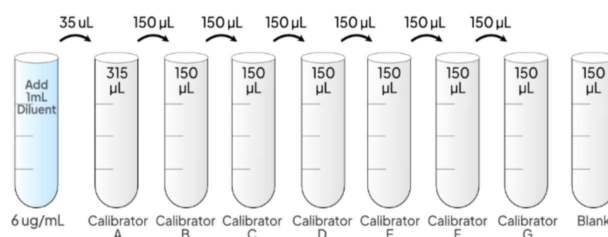
Prepare 1X Wash Buffer by diluting Wash Buffer 10X with deionized water.
To make 25 mL of 1X Wash Buffer, combine 2.5 mL Wash Buffer 10X with 22.5 mL deionized water. Mix thoroughly and gently.

Antibody Cocktail:

Prepare Antibody Cocktail by diluting the capture and detection antibodies in Diluent.
To make 500 µL of the Antibody Cocktail, combine 5 µL of Capture Antibody 100X and 5 µL of Detector Antibody 100X with 490 µL of Sample Diluent. Mix thoroughly and gently.

HSA Standard:

Transfer each lyophilized bead into a new tube. Resuspend each HSA lyophilized standard in 1 mL of Diluent, to get a stock concentration of 200 ng/mL. Dilute the stock HSA 10-fold to prepare a 20 ng/mL standard (Calibrator A). Store the rest of the reconstituted HSA standards at -20∘C.



	Concentration (ng / mL)	Preparation
HSA Standard	200	Resuspend in 1 ml of Diluent
Calibrator A	20	35 µL of HSA Standard to 315 µL of Diluent
Calibrator B	10	150 µL of Calibrator A to 150 µL of Diluent
Calibrator C	5	150 µL of Calibrator B to 150 µL of Diluent
Calibrator D	2.5	150 µL of Calibrator C to 150 µL of Diluent
Calibrator E	1.25	150 µL of Calibrator D to 150 µL of Diluent
Calibrator F	0.625	150 µL of Calibrator E to 150 µL of Diluent
Calibrator G	0.3125	150 µL of Calibrator F to 150 µL of Diluent
Blank	0	150 µL of Diluent

Sample preparation:

Prepare 75 µL of each sample to be tested (150 µL for duplicates). Dilute samples at least 2X using Diluent.

Typical Sample Dynamic Range	
Assay Range (O.D)	0.03 – 3.0
Sample Type	**Range**
Plasma	10MX – 320MX
Serum	5MX – 80MX
Urine	1kX – 20kX
CSF	40kX – 320kX

Note: Spin samples at 2,500 g for 10 minutes to remove platelets and debris.

Assay procedure:

1. Prepare all reagents, standards, and samples as directed above.
2. Remove excess microplate strips from the plate frame.
   - Seal and return them to 4° C storage.
   - The remaining strips should be used within two weeks if kept sealed.
3. Add 50 µL of each sample or standard to separate wells in the microplate strips.
4. Add 50 µL of the Antibody Cocktail to each well.
5. Seal the plate and incubate for 1 hour at room temperature on a plate shaker (400 rpm). Protect the plate from light during incubation.
6. Wash each well four times:
   - For the first wash, add 200 µL 1X Wash Buffer to the wells containing sample.
   - Aspirate each well.
   - Dispense 300 µL 1X Wash Buffer. Wash buffer should remain in the wells for at least 10 seconds.
   - Aspirate each well. Ensure complete removal of liquid between washes.
   - Repeat four times for a total of 4 washes. Use new tips for each washing step.
   - After the final wash, blot the plate against clean paper towels.
7. Add 100 µL of TMB Substrate Solution to each well. Incubate for 5 minutes in the dark on a plate shaker at 400 rpm.
8. Add 100 µL of Stop Solution to each well. Shake for 1 minute at 300 rpm. The solution will change from blue to yellow.
9. Measure the absorbance at 450 nm (endpoint reading). If wavelength correction is available, set to 620 nm.

Data analysis:

We recommend running the samples and calibrators in duplicate. To calculate the results, average the duplicate OD readings and subtract the average OD of the zero standard. Fit the standard curve using 4PL or a linear regression on a log-log plot. Interpolate the standard curve to determine the samples' concentrations. Multiply the calculated concentration by the dilution factor to obtain the final concentration. A new standard curve should be generated with every experiment.

Warnings and precautions:

The Diluent contains ProClin 300®, which may cause an allergic skin reaction. Avoid ingestion and contact with eyes, skin, and mucous membranes.

Example of a standard curve: